



Novel MALDI Imaging solution empowered by a timsTOF fleX and dedicated bioinformatics pipeline for identification of lipids from tissue

MALDI Imaging spectra can be used to investigate how compounds are localized across tissue samples. In this application note, we present a novel software workflow for the identification of signals using data from the high-speed, high spatial resolution timsTOF fleX instrument.

Introduction

Understanding the content and distribution of molecules in tissue and the changes that occur to them in disease is integral to enhancing our understanding of the tissue-based diseases, such as cancers and neurologicalbased conditions. MALDI Imaging is a powerful label-free technique that allows mapping of a wide range of molecules from tissue sections and is capable of detecting hundreds to thousands of unique compounds in a single sample. This allows the understanding of molecular distributions over entire samples or even to classify distinct regions of interest. We present a novel workflow solution for automatically annotating MALDI Imaging data from a timsTOF fleX, allowing

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Figure 1: Automated annotation of metabolites and lipids from MALDI Imaging data workflow. *Lipid Maps and HMDB are not Bruker products.

subsequent generation of annotated images. In this application note, the workflow above is demonstrated on measurements of lipids from rat brain sections.

Methods

Fresh-frozen tissue sections on conductive glass slides were dried then matrix coated using a TM Sprayer (HTX Technologies, Chapel HIII, NC, USA). MS data were acquired on a timsTOF fleX. The measurement parameters were: positive mode, m/z range 300-1000, 20 µm pixel size, 10 kHz laser repetition rate. Data were visualized using SCiLS Lab (Bruker Daltonik GmbH). MetaboScape (Bruker Daltonik GmbH) was used for lipid annotation using the customizable Analyte List tool to match against the

LIPID MAPS structure database and lipids reported in literature. MS data were processed in MetaboScape using the T-ReX² algorithm for feature extraction, de-isotoping and ion deconvolution. 2500 total pixels were averaged in 5x5 blocks yielding 100 spectra with averaged signal intensities. Mass tolerances were set to 5 mDa, and 250 mSigma for isotopic pattern fits.

	m/z meas.	M meas.	lons	Name	mSigma	∆m/z [ppm]	Molecular For	An	AQ 🔻
1	788.61566	787.60839	± •	PC(18:0/18:1(11Z))	14.2	-0.978	C44H86NO8P	AL.	11
2	369.35237	368.34509	<u>+</u> •	3-Deoxyvitamin D3	20.5	2.060	C ₂₇ H ₄₄	AL)	
3	744.49387	743.48659	<u>+</u> •	LMGP02080001	20.4	-3.129	C43H70NO7P	AL.	
4	772.52494	771.51766	± •	LMGP02080002	36.8	-3.426	C45H74NO7P	AL)	
5	734.56932	733.56204	± •	PC(10:0/22:0)	15.2	-0.172	C40H80NO8P	AL	11
6	706.53835	705.53108	± •	PC(10:0/20:0)	8.1	-0.279	C38H76NO8P	AL.	11
7	651.53501	650.52773	± •	LMGL02070009	16.8	0.450	C43H70O4	AL	11
8	496.34015	495.33288	± •	PC(O-14:0/2:0)	15.7	0.781	C24H50NO7P	AL.	11
9	746.60565	745.59838	± •	PC(O-16:0/18:1(9Z))	20.5	-0.680	C42H84NO7P	AL.	11
10	792.55377	791.54649	± •	PC(15:0/22:6(4Z,7Z,10	17.3	-0.669	C45H78NO8P	AL)	11

Table 1: Automatically annotated compounds using MetaboScape 5.0. Using a custom Analyte List of known compounds the features (de-isotoped m/z signals belonging to the same molecular formula) extracted by the T-ReX² algorithm were automatically annotated. The entries of the Analyte List, containing 42,755 compounds were downloaded from Lipid Maps (http://www.lipidmaps.org). Annotation Quality Scoring provided a fast overview of the confidence of each annotation based on accurate mass and isotopic fidelity (AQ column). Table 1 shows 10 hits out of 47 automatically annotated features.

Results and Discussion

Label-free MALDI Imaging is capable of demonstrating localization of more molecules and molecular classes in a single tissue section than immunohistochemistry or other tag-based imaging techniques. A single MALDI Imaging dataset may contain hundreds to thousands of unique, label-free ion images, which can be used for molecular marker discovery or investigating the molecular content of specific regions of interest. Particularly for SpatialOMx studies, MALDI Imaging provides morphological topography to a given sample. For timsTOF fleX MALDI Imaging datasets, we present a pipeline for compound identification and visualization (Figure 1). Using this, timsTOF fleX peak list data is loaded into SCiLS Lab (version 2020a) for statistical

analysis and defining regions of interest (ROIs). The spectral and ROI data is loaded into MetaboScape (version 5.0) for annotation e.g. via databases. The annotated data can be exported back to SCiLS Lab for visualization.

Table 1 is an example of a MetaboScape 5.0 data output from a timsTOF fleX MALDI Imaging dataset. The first two column headings indicate features, i.e. the de-isotoped m/z signals belonging to the same molecule extracted by the T-ReX² algorithm. The Ions column shows which ion adducts are combined into a feature (e.g. M+H). mSigma indicates goodness of fit of the isotopic pattern (measured vs simulated), where a low value denotes better fit. $\Delta m/z$ [ppm] specifies mass error of measured versus predicted mass in

parts per million. Annotation specifies how the annotation was completed (e.g. database or analyte list), and AQ (Annotation Quality) provides a fast overview of the scoring based on accurate mass and isotopic fidelity matching based on user-defined settings.

Figure 2 shows the distribution of m/z 788.623 in rat brain, identified as PC(36:1) in MetaboScape with the annotation in SCiLS Lab. As a comparison of the quality of the annotation and distribution, the right panel shows the distribution of PC(36:1) previously identified from published literature. In both the timsTOF fleX and published data, the signal is located in the white matter tracts of the brain.



Figure 2: Lipid distribution. Visualizaiton of PC(36:1) in the dataset used for compound annotation with MetaboScape. The lipid distribution could be confirmed with a reference from literature. Scale bar represents 1 mm.

Conclusions

- Lipid annotation automated with MetaboScape 5.0 featuring T-ReX² feature extraction technology and AQ scoring. Visualization of annotated signals with SCiLS Lab completing the intuitive workflow.
- Lipid distributions map to the expected localizations.
- This workflow enables SpatialOMx by providing annotations to the morphological topography.

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